## **Variation in the Physico-Chemical Properties of Badagry and Ikorodu Soils, Lagos Nigeria**

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## **Abstract**

*This study examined the variation in the physico-chemical properties of soils in Badagry and Ikorodu, Lagos to establish the effect of season and location on soil physical and chemical properties. Soil samples were taken at depths of 0-20cm from 26 and 36 points respectively at Badagry and Ikorodu using soil auger and collected in polythene bags The soil samples were analyzed for their texture, structure, pH, and the availability of some basic soil nutrients such as Nitrogen, Organic Carbon, Potassium, Phosphorus, etc) in accordance with Standard analytical procedures. The study revealed that the physico-chemical properties of soil in the areas under focus do not significantly vary among the variables (location, season and vegetation cover) probably because of the similarities in geology, climate and vegetation types..* 

## *1.0 Introduction*

The main threats to soils are increasing urban areas, road building and industrial development, erosion, acidification, accumulation of pollutants, organic matter loss and deteriorating soil structure (Aweto, A.O. and Ekuigbo U.E, 1994). Soil contamination by heavy metals can originate from a number of sources including geological parent material, industrial processes (atmospheric emission, waste disposal, and effluent disposal) and farming practices. Contaminants usually seep down into the soil and even remain long after the contaminant has left the surface of the soil. Land use pattern has also had a significant impact on the quality of the soil in a typical environment.

The maintenance of natural systems or soil fertility in tropical forest ecosystems is achieved by high and rapid circulation of nutrients through the fall and decomposition of litter which is a function of the season. The decomposed litter is also the basis of many food chains in tropical forests and is a principal source of energy for the biota of the forest floor and soil, where the trophic chain of detritus predominates (Spain, 1984; Ola-Adams and Egunjobi, 1992; Oliveira and Lacerola, 1993; Regina *et al*., 1999). Decomposition is a key process in the control of nutrient cycling and formation of soil organic matter (Berge B. and McClaugherty, 2002).

There is abundant literature, in the humid tropics, on soils physico-chemical and biological changes following deforestation and subsequent land cultivation (Ghuman and Lal, 1991; Juo *et al*, 1995; San José and Montes, 2001; Koutika *et al*., 2002; Schroth. *et al*., 2002; Whitbread. *et al.,* 2003; Sisti *et al*., 2004; Tchienkoua and Zech, 2004; Walker and Desanker, 2004). Extensive work has been done on the conversion of natural forests into agroforests and cultivated land systems (Lal, 2001; Walker and Desanker, 2004), as well as with soil organic matter dynamics in African tropical forests (Moyo, 1998; Rishirumuhirwa and Roose, 1998; Walker and Desanker, 2004). This study, however, examines the seasonal variations in the physicochemical properties of soil in two distinct ecosystems, the coastal/hydromorphic soil in Badagry and the upland soil in Ikorodu area of Lagos. In order to establish the prevailing quality of the soil in the study area, soil samples were collected and analysed. Soil samples were collected from two distinct locations (Badagry and Ikorodu) using a Garmin Global Positioning

For the purpose of this paper, soil sample were taken during the two distinct prevailing seasons in the country, the wet and dry season, so as to assess the effect of the prevailing weather condition to the concentration of soil nutrients and the physical state of the soil.

System (GPS) 12 XL™.

Lagos, the most populated state in the country was selected as the area of study. However, the study was further streamlined to two major areas in Lagos, namely Badagry and Ikorodu local government areas.

Two main vegetation types are identifiable in Lagos State: Swamp Forest of the coastal belt and dry lowland rain forest. The swamp forests in the state are a combination of mangrove forest and coastal vegetation developed under the brackish conditions of the coastal areas and the swamp of the freshwater lagoon and estuaries. Red mangrove (sometimes attaining heights of 592m) as well as mangrove shrubs, stilt rooted trees with dense undergrowths and raffia and climbing palms are characteristic of the swamp forest zone. Of course, on the seaward side of this zone, wide stretches of sand and beaches exist. Although a small amount of pit props and fuel material emanate from the swamp forest zone in Lagos State, it is of no significance in the lumber economy of Nigeria.

Lying to the north of the swamp forests is the lowland (tropical) rain forest zone. This zone, which stretches from the west of Ikeja through Ikorodu to an area slightly north of Epe, has been modified by man. Yet this is the area of the state where such economically valuable trees as Teak (*tectona grandis*), *tripochiton*, *seletrocylon* (Arere), *Banclea diderrichil* (Opepe) and *Terminahia* (Idigbo) are to be found. The creeks, lagoons and rivers act as arteries which carry huge quantities of logs from out of state sources to Lagos.

Lagos State is endowed with very little arable land. Altogether, four soil groups are identifiable. On the western half of the coastal margin, juvenile soils on recent windborne sands occur. The rest of the coastal area towards the east is covered also by juvenile soils on fluviomarine alluvium (mangrove swamp).

Thirdly, a narrow and rather discontinuous band of mineral and/or organic hydromorphic soils occurs in the middle and northern-eastern sections of the state. The fourth group, occurring in two rather tiny and discontinuous patches along the northern limits of the state, consists dominantly of red ferrallitic soils on loose sandy sediments. Specifically, the study areas selected in Lagos lie between 6°46'N and 2°23"E with an elevation of about 19ft at Badagry and 6°36"N and 3°00"E with an elevation of about 47ft at Ikorodu. Badagry falls into the ecological zone of wetland soils and lies on the coast where inland water empties into the Atlantic Ocean. It has a geologic origin of deltaic basis and tidal flats (FADAMA, 2011). The natural vegetation is mangrove. The floras of the area consist of *Rhizophora* mangle and *Rhizophora racemosa* (otherwise referred to as red mangrove and black mangrove respectively). These two species are strongly zoned, with the former occupying areas closer to the water while the latter are in the upper reaches. Other species occurring to a lesser extent include *Avicennia Africana*, *Laguncularia racemosa*; plus the palms *Prodococcu wateri* and *Ancistrophylum opacum*.

Ikorodu on the other hand, has some parts falling into wetland zone and its other part falling into rainforest ecological zone. Its soils developed from recent alluvium and coastal plain sands (FADAMA, 2011). It is a forested area in which very tall trees abound. Some of the common trees are *Afromosia Laxiflora, Burkea africana, Daniella oliveri and Laoberlinia doka*.

The study areas exhibit similar climatic conditions. They are characterised by a humid tropical climate characterized by distinct dry and wet seasons with moderate mean annual rainfall which varies between 1381.7 mm and 2733.4 mm in recent time from one location to the other. However, the average rainfall across Lagos for over 25 years is estimated at about 2,500mm. There are two discernible seasons (rainy and dry seasons) but there is hardly a month without precipitation in Lagos. A double maxima of rainfall regime are recognizable from March to early July and the other from September to early November with a break in late July and August. The maximum temperature ranges between  $29^{\circ}C - 34^{\circ}C$ , the lowest being in the month of July and the highest in February. The minimum temperature varies between  $24^{\circ}$ C -  $28^{\circ}$ C. The relative humidity is generally high and rarely below 70 % throughout the year. During the wet season months, the south west winds prevail as the front moves to the north. But as from October when the front moves south wards, the northeast winds sweep in the dry season. Lagos State, however, experiences predominantly south-westerly wind and sea breezes all year round.

# Map of southwest Nigeria showing the study area (Lagos State)



**Fig 1: Map of Southwest Nigeria showing the study area (Lagos State)**

## **Lagos State**

**Fig2 :** *showing Administrative Map of Lagos State*





**Fig 3: Map showing sampling points**



**Fig 4: Map showing sampling points**

#### **1.1 Sampling and Analytical Procedure**

A total of 27 and 36 soil samples were taken at Badagry and Ikorodu respectively at depth of 0-20cm, with the aid of a soil auger. 9 soil samples each were collected during the dry and wet seasons and 9 soil sample each were collected from cleared areas and forested areas at Ikorodu. 7 soil samples were collected during the dry season and 6 soil samples during the rainy season at Badagry and 7 soil samples each from cleared areas and forested areas at the same location. These samples were collected in polythene bags and transferred to the laboratory for analysis. The soil samples were analyzed in accordance with Standard analytical procedures (British Standards [BS] and American Society for Data Testing)

For soil textural analysis, soil sample collected was subjected to mechanical analysis for particle size and soil textural classification. The mechanical analysis was carried out on the soil samples by the Bouyoucos method to determine the various sizes of particles present in the fine earth (i.e. particle  $\langle 2mm \rangle$ ) of the soil using international scale of: 50g of the air-dried 2mm sieved soil were placed into container of a high-speed stirrer. This was followed by the addition of 25ml of 5% calgon and stirred with high speed for 15 minutes. The content of the container was then transferred to a 1 litre cylinder (tall form), diluted to mark and stirred for one minute with a wooden paddle. This was followed by inserting a Bouyoucos soil hydrometer for 20 seconds before reading the International Silt and Clay (<20 $\mu$ ) after 4 minutes, 48 seconds and International Clay (<2 $\mu$ ) after 5 hours.

The temperature of the suspension was taken after each reading and 0.3 units added or (subtracted) for every degree above (or below) 19.50C. After the second hydrometer reading, most of the suspension was decanted, refilled with water, paddled and allowed settling for 4 minutes 48 seconds before decanting again. This procedure was repeated until the supernatant liquid was clear. The sand residue was then transferred to a weighed porcelain basin and weighed again to obtain the weights of sand (coarse + fine sand), silt and clay percentages in the 50g soil were then calculated. On the other hand, for Chemical Properties, the pH values of the soil samples were determined in the laboratory using a HH4 Ionoscope pH meter. The pH was determined by pouring 1:2.5 soil water suspensions that had been stirred and allowed to equilibrate for about 1 hour into the electrode.

For Exchangeable cation, 2.5g portions of finely ground representative samples were shaken in a conical flask with 25ml of 1N ammonium acetate for about 1 hour and filtered into plastic cups. The filtrate was used for the determination of sodium (Na+), Potassium (K+), Calcium (Ca<sup>2+</sup>) and Magnesium (Mg<sup>++</sup>), using a Flame photometer. The concentrations of the cations were calculated after due note of the dilution factors and expressed either in parts per million (ppm) or milligrams equivalent per 100g soil (meq/100g soil). Also, heavy metal content of the sample was determined using Perkin Elmer 2380 double beam Atomic Absorption Spectrophotometer while Carbon was determined by the wet combustion method of Walkey and Black (1934). 1g of finely ground representative sample was weighed in duplicate into each beaker and rotated gently to wet the soil sample completely. This was followed by the addition of 20ml of conc.  $H_2SO_4$  using a graduated cylinder, taking a few seconds only in the operation. The beaker was rotated again to effect complete oxidation and allowed standing for 10 minutes before dilution with distilled water to about 200-250mL. 25 ml of 0.5N ammonium sulphate was then added and titrated with 0.4N Potassium permanganate.

For total Nitrogen**,** 2.5g of a representative air dried soil were accurately weighed into Tecator digestion flasks and a catalyst mixture containing selenium, CuSO4 and Na2SO4 was added followed by 10ml of concentrated sulphuric acid. The contents of the flask were mixed by gentle swirling and then digested on a Tecator block until the digest became clear. Heating was continued for another one hour before the digest was allowed to cool. The digest was then transferred quantitatively with distilled water to a 150ml conical flask and made up to mark with distilled water. Aliquot of this was analysed and the percentage Nitrogen content of the soil was then calculated after taking into account, the different dilution factors. Also, available phosphorus in the soil sample was determined by weighing 1g of sample into an extraction flask. This was followed by the addition of 10ml of Bray P-1 extraction solution (0.25N HCl & 0.2N NH4F and shaking immediately for 1 minute and filtered. 5mL of the filtrate was then measured into 250ml volumetric flask and diluted to about 220ml with distilled water followed by 4ml of ascorbic acid solution (0.056g ascorbic acid in 250ml molybdate – tartarate solution) and diluted to mark. This was allowed to wait for at least 30 minutes for full colour development before reading from at 730nm and lastly electrical conductivity of the soil sample was determined on the filtrate obtained after filtering the suspension used for the pH determination.

## **1.2 Data Analysis**

In collecting the soil data, the experimental design used was a three-factor factorial experiment in completely randomised design. The three factors are location (Ikorodu and Badagry), season (rainy and dry), and vegetation cover (cleared and forested). Consequently, the analysis of variance (ANOVA) procedure was carried out to determine the variations that exist in the soil properties at the study sites. The analysis involved testing for significant differences between levels of each factor and the interactions between the factors. The General Linear Model analysis was done using Statistical Package for Social Scientists (SPSS).

#### **1.3 Results Of Analysis Of Soil Samples**

Detailed results of the data analysis are presented in Appendix 4 while a summary of the ANOVA table is shown below (Table 1-20).

<b>Source of Variation</b>	Degrees of freedom	Sum of <b>Squares</b>	<b>Mean Squares</b>	<b>F-calculated</b>	Sig.
Location $(L)$		0.095	0.095	$16.179*$	0.000
Season $(S)$		0.372	0.372	$63.483*$	0.000
Forest Type (FT)		0.022	0.022	3.739ns	0.058
<b>LxS</b>		0.001	0.001	0.219ns	0.642
LxFT		0.214	0.214	$36.448*$	0.000
SxFT		0.001	0.001	0.125ns	0.725
LxSxFT		0.000	0.000	0.058ns	0.810
Error	54	0.317	0.317	0.006ns	
Total	61	0.994			

**Table 1: Analysis of Variance table for Soil Reaction (Sand)**

*\* denotes "significant" at 0.05 level while ns denotes "not significant" at 0.05 level.* **Source:** *Field Data 2010*





*\* denotes "significant" at 0.05 level while ns denotes "not significant" at 0.05 level.* **Source:** *Field Data 2010*

**Table 3: Analysis of Variance table for Soil Reaction (Clay)**

<b>Source of Variation</b>	Degrees of freedom	Sum of <b>Squares</b>	<b>Mean Squares</b>	<b>F-calculated</b>	Sig.
Location $(L)$		0.026	0.026	9.612ns	0.003
Season $(S)$		0.095	0.095	35.391*	0.000
Forest Type (FT)		0.008	0.008	2.834ns	0.098
<b>LxS</b>		0.000	0.000	0.091ns	0.764
LxFT		0.010	0.010	3.808ns	0.056
<b>SxFT</b>		0.001	0.001	0.359ns	0.552
LxSxFT		0.001	0.001	0.218ns	0.166
Error	54	0.145	0.003		
Total	61	0.293			



#### **Table 4: Analysis of Variance table for Soil Reaction (Organic Carbon)**

*\* denotes "significant" at 0.05 level while ns denotes "not significant" at 0.05 level.* **Source:** *Field Data 2010*





*\* denotes "significant" at 0.05 level while ns denotes "not significant" at 0.05 level.* **Source:** *Field Data 2010*





*\* denotes "significant" at 0.05 level while ns denotes "not significant" at 0.05 level.* **Source:** *Field Data 2010*













*\* denotes "significant" at 0.05 level while ns denotes "not significant" at 0.05 level.* **Source:** *Field Data 2010*





*\* denotes "significant" at 0.05 level while ns denotes "not significant" at 0.05 level.* **Source:** *Field Data 2010*













*\* denotes "significant" at 0.05 level while ns denotes "not significant" at 0.05 level.* **Source:** *Field Data 2010*





*\* denotes "significant" at 0.05 level while ns denotes "not significant" at 0.05 level.* **Source:** *Field Data 2010*













*\* denotes "significant" at 0.05 level while ns denotes "not significant" at 0.05 level.* **Source:** *Field Data 2010*





*\* denotes "significant" at 0.05 level while ns denotes "not significant" at 0.05 level.* **Source:** *Field Data 2010*









#### **1.4 Discussions and Conclussions**

From the Data analyzed, it is discovered that the parameters involved react differently to the main effects which are; location (Ikorodu and Badagry), seasons (wet and dry) and vegetation cover (cleared and forested).

## **1.4.1 Effect of Location**

Analysis of variance as shown in Table 1 reveals that that Location (L) is highly significant to sand i.e. there is significant difference in sand of Ikorodu and Badagry while Table 2 indicates that there are no significant differences in silt of Ikorodu and Badagry. For clay in Table 3, it shows that Location (L) is not significant, meaning that there is no significant different in clay of Ikorodu and Badagry. Organic carbon in Table 4 indicates Location (L) is not significant to organic carbon content of Ikorodu and Badagry while Table 5 also shows no significant difference in the Nitrogen content of the two sites. i.e. Ikorodu and Badagry. Table 6 also indicates that Location (L) is highly significant to Phosphorous (Bray II), ppm content of the two sites while Table 7 shows that Location (L) is not significant to Potassium level of the two sites. Table 8 revealed that there is no significant difference in soil moisture content of the two sites. For Table 9, it shows that there is significant difference in soil P<sup>H</sup> of Ikorodu and Badagry. Tables 10, 11, 12 and 13 indicates that there are no significant difference in conductivity, sodium mg/kg, calcium, and magnesium mg/100g of the two sites. Tables 14 and 15 show that there are significant differences in sulphate mg/100g and chloride, mg/100g at the two sites while Table 16 indicates that there are no significant differences to iron and manganese mg/100g of the two sites. Table 18 also indicates that there is significant difference in zinc, mg/100g of Ikorodu and Badagry respectively. Table 19 shows that Location (L) is not significant to lead, mg/100g of Ikorodu and Badagry while Table 20 indicates that Location (L) is not significant to Copper, mg/100g.

## **1.4.2 Seasons (Dry and Wet)**

Analysis of variance from Tables 1 and 2 indicate that there are no significant differences in sand and silt while Table 3 shows that Seasons (S) are significant, meaning that there is significant difference in clay of Ikorodu and Badagry. For Tables 4 and 5, it shows that Seasons (S) are highly significant to organic carbon and nitrogen content of the soil, while Tables 6 and 7 indicates that Seasons (S) are not significant to Phosphorous and Potassium level in the study areas. Table 8, shows that Seasons (S) are significant to soil moisture content of the two areas, while Table 9 indicates that Seasons (S) are not significant to  $P^H$  content of the study area. Meanwhile, Table 10 indicates that Seasons (S) are highly significant to conductivity of the study areas. Tables 11, 12, 13 and 14 shows that Seasons (S) are not significant to sodium, mg/100g, calcium, magnesium, mg/100g and sulphate, mg/100g. Table 15, indicates that Seasons (S) are not significant to chloride while Table 16, shows that Season is highly significant to iron. Table 17 indicates that Seasons (S) is not significant to manganese, mg/100g. for Tables 18 are highly significant to zinc mg/100g . Table 19 indicates that Season (S) are not significant to lead, mg/100g while Table 20 shows that Seasons (S) are highly significant i.e. there is significant difference in copper mg/100g, in the two seasons.

## **1.4.3 Vegetation Cover (Cleared and Forested)**

Analysis of variance from Tables 1, 2 and 3 show that there are no significant differences for sand, silt and clay in cleared land and forested land.

Tables 4, 5 and 6 all indicated that there are significant differences in organic carbon, nitrogen and phosphorous (Bray II) between the Forest Type (FT) of the study areas. Table 7 shows that Forest Type (FT) are not significant to Potassium level in the study areas, while Table 8 indicates that Forest Type (FT) are highly significant to moisture content of the study areas. Table 9,10,11,12,13,14,15, 16, `17, 18, 19 and 20 shows that Forest Types are not significant to  $P^H$ , conductivity, sodium mg/kg, calcium, magnesium, mg/100g, sulphate, mg/100g, chloride, mg/100g, iron, mg/100g, manganese mg/100g, , zinc mg/100g, lead mg/100g and copper mg/100g of the study areas.

## **1.5 Conclusion**

Results and discussion have revealed that the physico-chemical properties of soil in the areas under focus do not significantly vary among the variables. The non statistical variation in the physical and chemical properties of soil across location, season and vegetation covers could be attributed to the fact that the two area are under the same geological formation of sedimentary rock, under similar tropical climate and vegetation types with little micro climate differences , especially around the coastal settlement of Badagry Therefore, management intervention like, soil enriching intercropping or inter-rotational planting, manuring and composting, fertilizer application etc. would be useful.

## *References*

- **Adejuyigbe C. O.** (2000). *Effects of fallow legumes on soil micro arthropods and their roles in nutrient turnover under humid tropical conditions.* (PhD Thesis.) University of Ibadan, Ibadan, Nigeria.
- **Aerts R.** (1996). Nutrient resorption from senescing leaves of perennials; are there general patterns? *J. Ecol.* **84**: 597-608.
- Aiboni V. U. (2001). Characteristics and classification of soil of a representative topographical location in University of Agriculture, Abeokuta. Asset Series A **1**(1): 51–61.
- **Alfred E. H.** and **O'Sullivans J. N. O.** (2001). Leaf litter decomposition of *Piper aduncum, Gliricidia sepium*  and *Imperata cylindrical* in the humid lowland of Papau New Guinea. *Pl. Soil* **230**: 115–124.
- **Anderson J. M.** and **Swift M. J.** (1983). Decomposition in tropical forest. In *Tropical rainforest. Ecology and management.* (S. L. Sutton, T. C. Whitmore and A.C. Chadwick, ed.), pp.287-309. Blackwell, Oxford.
- **Aweto, A.O. and Ekuigbo U.E. (1994).** Effects of Oil Palm Plantations on Tropical Forest Soils in South Western Nigeria. *The Indonesia Journal of Geography 26,51-59*.
- **Berge B.** and **McClaugherty C.** (2002). *Plant litter decomposition humus*. Springer-Verlag, New York.
- **Black C. A** (ed.) (1965). *Methods of soil analysis*. Agronomy No. 9, Part 2. America Society of Agronomy, Madison, Wisconsin.
- **Boulton A. J.** and **Boon P. L.** (1991). A review of methodology used to measure leaf litter decomposition in lotic environment. Time to turn over an old leaf? *Aust. J. mar. Freshwat. Res*. **42**: 1–45. *West African Journal of Applied Ecology - Volume 13*
- **Burghouts T. B. A., Van Straalen** and **Bruignzeel L. A.** (1993). Spatial heterogeneity of element and litter turnover in a Bornean rown forest. *J. Trop. Ecol.* **14**: 477–506.
- **Brenner J. M.** (1965). Total Nitrogen. In *Methods of soil analysis* (C.A. Black, ed.) Part 2, America society of Anatomy, Medison.
- **Courteaux M. M., Bottner P.** and **Berg B.** (1995). Litter decomposition, climate and litter quality trend. *Ecol. Evol*. **10**: 63–66.
- **Cox P., Wilkinson P. M.** and **Anderson J. M.** (2001). Effects of fungal inocula on the decomposition of lignin and structural polysaccharides in *Pinus sylvestris* litter. *Biol. Fert. Soils* **33**: 246–251.
- **De Santo A. V., Berg B., Rutigiliano F. A., Aleani A**. and **Frioretto A.** (1993). Factors regulating early stage of decomposition of needle litters in five different coniferous forests. *Soil Biol. Biochem*. **25**: 1423–1433.
- **Egunjobi J. K.** (1971). Ecosystems processes in a stand of *Ulex europaeus* L. (11) recycling of chemical elements in the ecosystem. *J. Ecol*. **59** 669–678.
- **Egunjobi J. K.** and **Fasheun F. E.** (1972). Preliminary observations on the monthly litterfall and nutrient content of *Pinus caribea* leaf litter. *Niger. J. Sci*. 6(1).
- **Facelli J. M. and Pickett S. T. A.** (1991). Plant litter its dynamics and effects on plant community structure. *Bot. Rev.* **57**: 1– 33.
- **Frankland J. C.** (1992). Mechanisms in fungal succession. In *The fugal community: Its organization and role in the ecosystems,* 2nd edn. (G. C. Carroll and D. T Wicklow ed.), pp. 383–401. Marcel Dekker, New York.
- **Frioretto A., Musacchio G. Andolfi** and **De Santo A. V**. (1998). Decomposition dynamics of litters of various pine species in a Corsican pine forest. *Soil Biol. Biochem*. **30**: 721–727.
- **Hermansah A. Z., Tsugiyuki M.** and **Toshiyuki. W**. (2002). Litter fall and nutrient flux in tropical rain forest. *West Sumatra, Indonesia 17th WCSS, 14*–*21 Aug. 2002. Thailand. Symposium No. 1125. Paper No. 1125*.
- **Hobbie S. E.** (1996). Temperate and plant spp. control over litter decomposition in Alaska tundra. *Ecol. Monog*. **66**: 503–522 (ISI).
- **Isaac R. A.** and **Kerber J. D.** (1972). Atomic absorption flame photometric techniques and uses in soil, plant and water analysis. In *Instrumental methods for analysis of soil and plant tissues.* (L. M. Walsh, ed.) Soil Science Society of America, Medison, Wilsconson.
- **Jackson M. C.** (1962). *Soil chemical analysis.* Prentic Hall, New York.
- **Kava'ova M.** and **Acek S. V.** (2003). Mountain Norway spruce forests. Needle supply and its nutrient content. *J. For Sci.* **49** (7): 327–332.
- **Keay R. W. J.** (1953). *An outline of Nigerian vegetation,* 2nd edn. Government printer, Lagos, Nigeria.
- **Klinge H**. and **Rodrigue W. A.** (1968a). Litter production in an area of Amazonian terra-firme forest. II. Mineral nutrient content of the litter. *Amazoniana* **1**(4): 303-310.
- Lisanework N. and Michelsen A. (1994). Litter fall and nutrient release by decomposition in three plantations compared with natural forest in the Ethiopian highland. *For. Ecol. Mgmt* **65**:149–164.
- **McClaugherty C.** and **Berg B.** (1987). Cellulose, lignin and nitrogen concentration as rate regulating factors in late stages of forest litter decomposition. *Pedobiologia* **30**: 101–112.
- **Muoghalu J. I., Adeloye O. M.** and **Balogun R. T.** (1994). Litter decomposition and inorganic element dynamics in a secondary rainforest at Ile-Ife. Nigeria*. Afr. J. Ecol*. **32**: 208–221.
- **Muoghalu J. I., Akanni S. O.** and **Eretan O. O.** (1993). Litter and nutrient dynamics in a Nigerian rainforest seven years after a ground fire*. J. Veg. Sci*. **4**: 325–328.
- **Nwoboshi L. C**. (1981). Nutrient cycling in managed teak plantation ecosystem. II. Litter fall and macro nutrient return to the forest floor. *Niger. J. agric. Sci.* **3**(1): 9–14.
- **O'Connel** (1988). Nutrient dynamics in decomposing litter in Karri *(Eucalyptus diversicolor* F. Muell) forests of South Western Australia*. J. Ecol*. **78**: 1186–1203
- **Ola-Adams B. A.** and **Egunjobi J. K.** (1992). Effects of spacing on litter fall and nutrient content in stands of *Tectona grandis* Linn. F. and *Terminalia superba* Engl. and Diels. *Afr. J. Ecol.* **30**:18–32.
- **Oliveira R.** and **Lacerola L. D**. (1993). Produ e compsiqu. Ca da serappilheira na Floresta da Tijula (Rz). *Revta brasil Bot*. **16**: 93–99.
- **Olson J. S.** (1963). Energy storage and the balance of producers and decomposers on ecological systems. *Ecology*  **44**:322–331.
- **Oohara H. N. and Yoshida N. K.** (1971). Balance of producers and decomposers in a grassland system in Obihiro. *J. Jap. Grassl. Sci*. **17**: 7–18.
- **Palm C. A.** and **Sanchez P. A.** (1990). Decomposition and nutrient release patterns of the leaves of three tropical legumes. *Biotropica* **22**: 330–338.
- **Regina M., Wetrington Braz Delitt** and **Vana stru FF aldi-De Vuono** (1999). Litter and nutrient content in two Brazilian tropical Forest. *Revta brasil. Bot.* **22**: 1999.
- **Reich P. B., Walter M. B.** and **Ellsworth D. S.** (1992). Leaf life-span in relation to leaf, plant and stand characteristics among diverse ecosystems. *Ecol*. *Monogr.* **62**: 365–392.
- **Reinaldo I. B.** and **Philip M. F.** (1995). *Carbon and nutrient flows in an Amazonian forest. Fine litter production and composition at Apiau*. Roraima, Brazil. *West African Journal of Applied Ecology - Volume 13*
- **Salamanca E. F., Kaneko N., Katagiri S**. and **Nagayama Y.** (1995). Effect of leaf litter mixtures on the decomposition of *Pinus densiflora* and *Quercus serrata* using field and laboratory microcosm methods*. XXth International Union of Forestry Research Organisation (IUFRO) World Congress Tampere, Finland, Aug. 6*–*12, 1995.*
- **Scott N. D.** and **Binkley D.** (1997). Foliage litter quality and annual net N mineralization; Comparison across North America Forest sites. *Oecoligia* **111**:151–159.
- **Sharma E.** and **Ambasht R. S.** (1987). Litterfall, decomposition and nutrient release in an age sequence of *Alnus nepalensis* plantation in the Eastern Himalaya. *J. Ecol*.**75**: 997–1010.
- **Singh J.** (1980). *Studies on structural and functional aspects of two subtropical humid forest type in Meghalaya.*  (PhD Thesis.) North-Eastern Hill University, Shillong, India.
- **Spain A. V.** (1984). Litter fall and the standing crop of litter in three tropical Australia rain forests. *J. Ecol.* **72**: 947–961.
- **Swift M. J., Heal O. W.** and **Anderson J. M.** (1979). *Decomposition in terrestrial ecosystems.* Blackwell, Stafford, UK. 372 pp.
- **Temel S.** (2003). Litter decomposition of *Picea orientalis, Pinus sylvestris* and *Castanea sativa* trees crown in Artvin in relation to their iInitial title quality variables. *Turkey Agric. For*. **27**: 23–243.
- **Swift M. J.** and **Anderson J. M.** (1989). Decomposition. In *Tropical Rainforest Ecosystems. Ecosystems of the World.* (H. Lieth and M. S. A. Weryer, ed.), pp. 547–569. Elsevier, Amsterdam.
- **Terrell T., Baker I. J., Graeme Lockaby B., William H. C., Calvin E., Meier John S. A.** and **Marianne K. Berke** (2001). Leaf litter decomposition and nutrient dynamics in four southern forested flood plain communities. *Soil Soc. Am. J.* **5**: 1334–1347.
- **Thorburn P. J., Probert M. E.** and **Robertson F. A.** (2001). Modelling decomposition of sugar cane surface residues with APSIM residue. *Fld Crops Res*. **70**: 223–232.
- **Tian G., Adejuyigbe C. O., Adeoye G. O.** and **Kang B. T.** (1998). Role of soil microarthropods in leaf decomposition and N release under various land-use practices in the humid tropics. *Pedobiologia* **42**: 33– 42.
- **Vanlauwe B., Diels J., Songinga N.** and Merckx R. (1997). Residue quality and decomposition: an unsteady relationship. In *Driven by native plant litter quality and decomposition*. (G. Cadisch and K. E. Galler, ed.), pp. 157–166.
- **Vituosek P.** and **Sanfiord R. I. (**1986). Nutrient cycling in moist tropical forest. *A. Rev. Ecol. System*.**17**: 1137– 167.
- **Walkley, A. and Black, I. A** (1934)., An examination of Degtjareff method for determining soil organic matter, and a proposed modification of the chromic acid titration method. Soil Sci., 1934, 37, 29–38